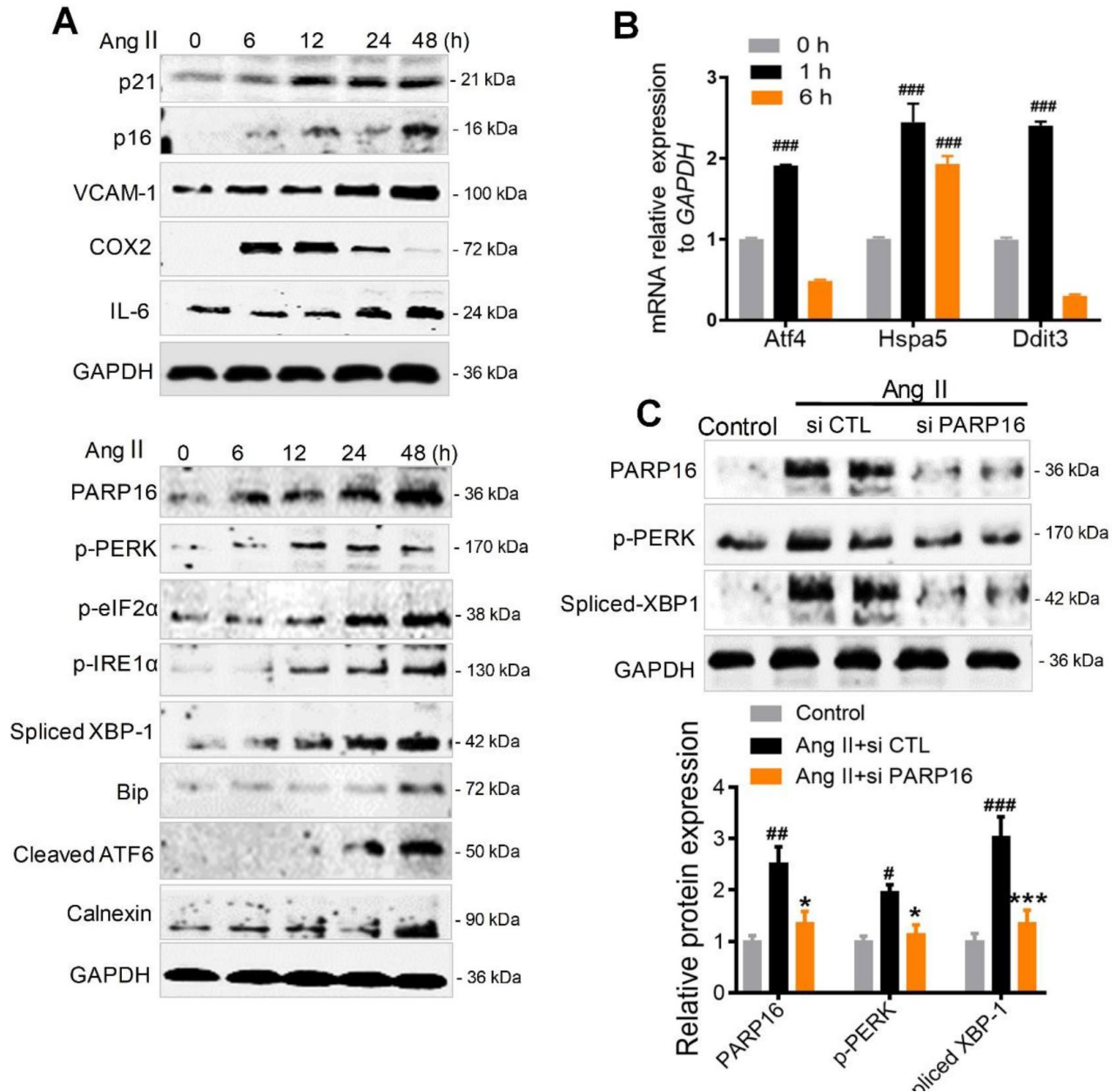
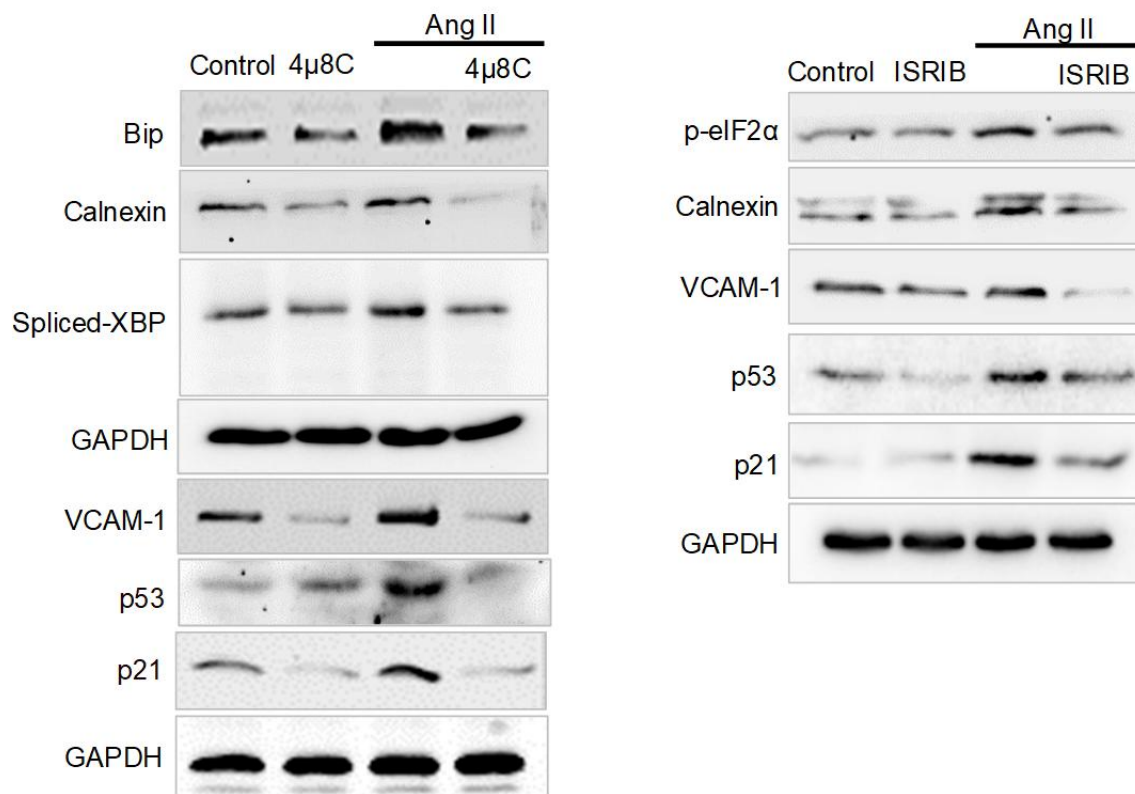


VSMCs

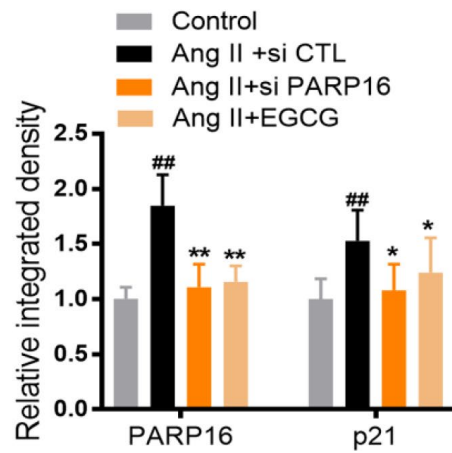
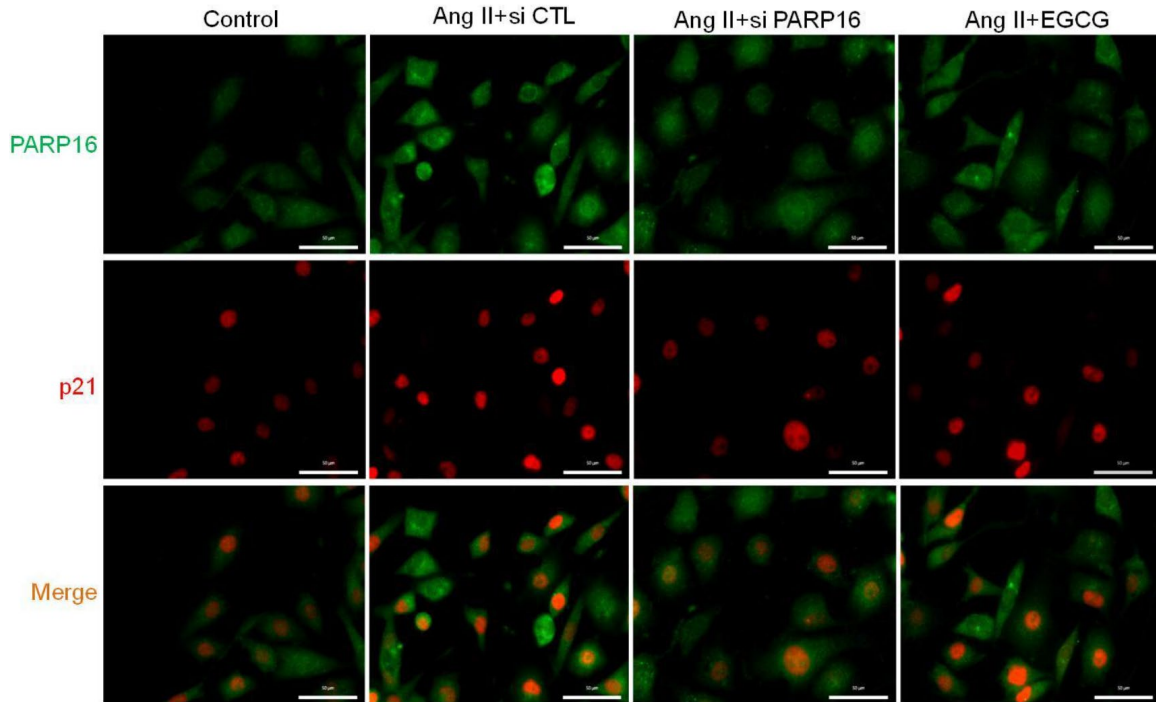


Supplementary Figure 1. PARP16 is involved in Ang II-induced VSMCs and HUVECs ER stress and senescence. (A) Senescence-associated markers, ER markers and PARP16 expression are upregulated after Ang II stimulation in VSMCs. At 0, 6, 12, 24 and 48 h after Ang II (2 μ M) administration, cells extracts were collected for determining the protein levels. (B) the UPR target genes (*Atf4*, *Hspa5*, *Ddit3*) is increased by Ang II treatment in VSMCs. (C) p-PERK, spliced XBP-1 were assayed by Western blot for VSMCs transfected with scramble (si CTL) or PARP16 siRNA before and after Ang II induction. Protein quantitative analysis was shown at the bottom of each Western blot. GAPDH serves as internal control, # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$ vs. control; * $p < 0.05$, *** $p < 0.001$ vs. Ang II+siCTL treated cells; all data were shown as mean \pm S.D. of at least four different replicates.



Supplementary Figure 2. Inhibitor of IRE1 or PERK signaling decreases the expressions of Ang II-induced RAECs senescence markers. RAECs were treated with Ang II (2 μM) for 48 h in the presence of 4μ8C (50 μM), ISRIB (1 μM) or DMSO vehicle, Bip, Calnexin, Spliced-XBP-1, p-eIF2α, p53, p21 and VCAM-1 levels were detected by Western Blot, respectively. Data shown are represent data from three independent experiments.

HUVECs



Supplementary Figure 3. Immunofluorescence double staining of PARP16 and p21 for HUVEC cells transfected with PARP16 siRNA or treated with PARP16 inhibitor (EGCG) before and after Ang II induction. Data were shown as mean \pm S.D of at least four different replicates; ^{##} $p < 0.01$ vs. control; ^{*} $p < 0.05$, ^{**} $p < 0.01$ vs. Ang II+siCTL treated cells.

PARP16 gene TSS upstream 2000bp

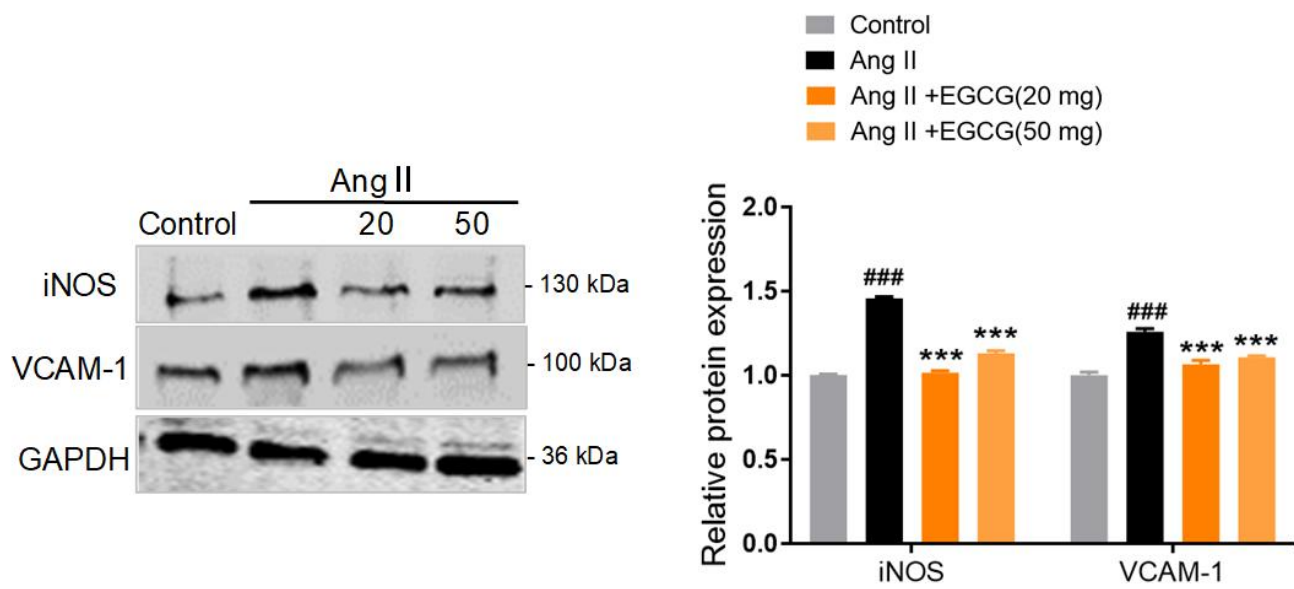
>rn6_refGene_NM_001014093 range=chr8:70710780-70712779 5'pad=0 3'pad=0 strand=+

repeatMasking=none

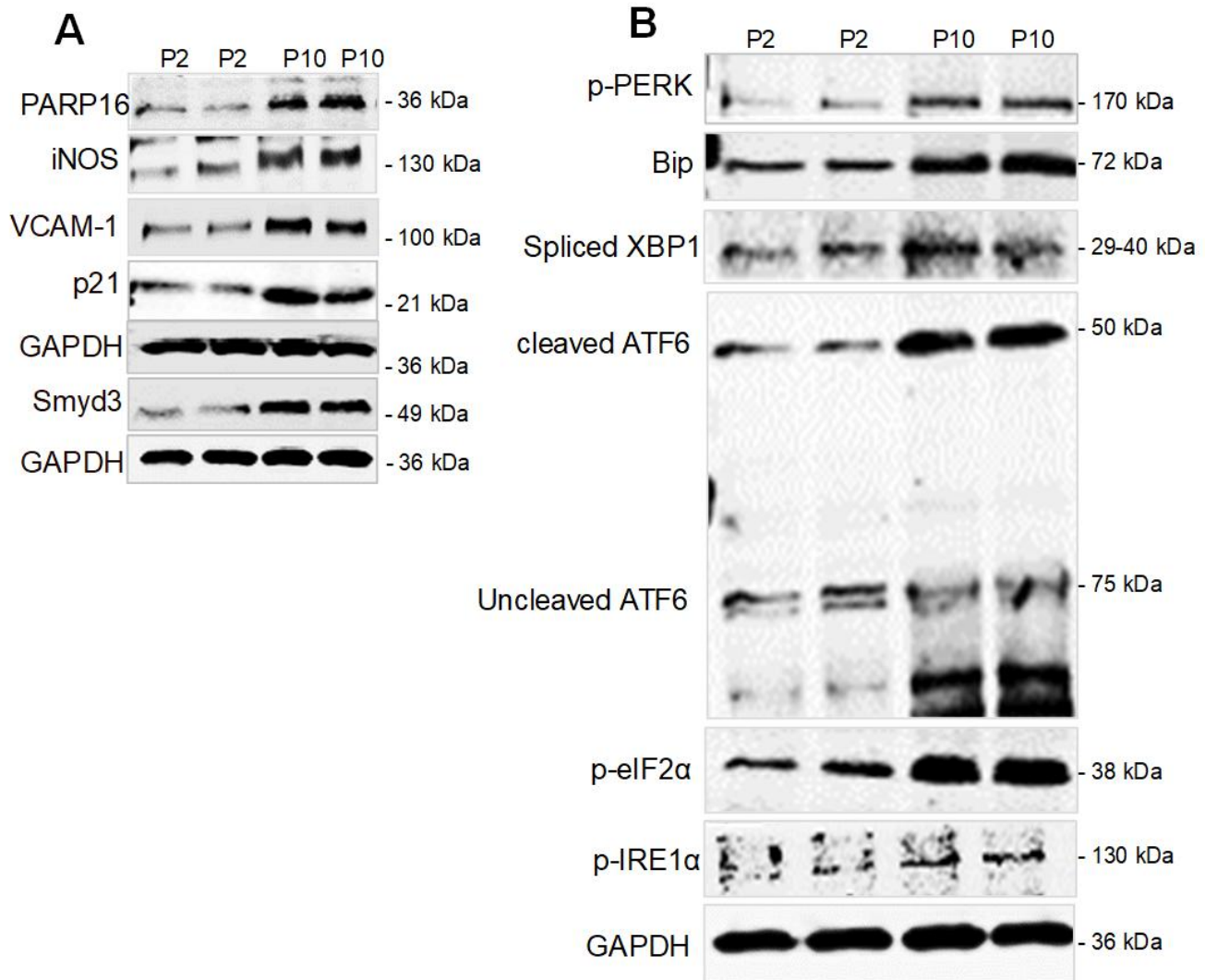
TATATAGTGAGAACTTTTCTAAGCAACAATAAACCAAAACCAACCAAAACCAACCAATAACAAC
CAACTAACCAAAACAAACAAGAAACCCCTACTGTTATGCCATTACTGTGGGTATACTCCCTGCCAG
TCATGGTGAGCATACCCCTGCAGTCACTGTGGATGTACCCCTTTCTGGGTGATAAGAGTGCCAA
CTTCAGAGGCTCTGAAGGGATGAGATGGACTCGATT**CCCTC**TACTGACATTACTAAATAACAAA
CAAGCAGACAGTAATAACACACCCCGAAATGGGAGGACAAGCAATAGTCTATCCTCTAAAATG
CCCTTTCTCTAGAACAACAGAAAT**GAGGG**CACAGAAGCAGAATAGGACTCTCTGTGAAGAGCA
AACAGAAGGGGTAGCAGGACTGACTGCAGAGTGCAGAGGTCCTTATGAATAAGGAGC
CAAAAGAGTATGATTA**GGAGGG**CAGGACTGCCAAAGGAGAATGTACATCAAATACAGCATA
TCACCGAAAGAAAGAACTGTGTTGTTTGTCTGAGTGCAGCATGGGCTACAGAGTAAGACCAC
AGAGAAAATAAGGGAAAGATTCCATACATTTTGTTCGTTGTTGCCAGTTGTTGTGGCACTGTG
GCATGGAGAATGCCCTTTACTTCCAGCACAAAAACAAACAAACAAACAAACAAACAAACGGCGAC
AAAAATAAGTCTTCTAGGTTTCCAGGAAGCCCAATGAACTTCAAGTAGCATCAATGCAAAAGTTT
ATAAAGAGAAACAACACGGCAAAGATATGGAGAGTCAAAGACCA**GGAGGG**AATCTTTTAAATTT
AAATTTTATTATTATCTTTTATTTGTTTGTGTTTCAACATAGGGTAGCCCTGGCTGTCCCTAGAATT
TGCTCTGTAGATCAGGCTGGCCTCAAACCCACAGAGATCAACCTGTCTCTGCCCTCTGAGTACT
GGATGAATAAATTTAAAGATTTATTTCTTAATTTTTTTTTTTGGTGAAGTTGTGTTTGGG
ATTTTGTTCGTTTTTTGAGACAGGGTTTCTCTGTGTAGCCTTGGCTGTCCCTGGAACCTGAT
CTGTAGACCAGTCGGGACTTGAACACAGAGATCTGCCTGCCCTGCTGGGACTAAAAGTGTGT
CATATGTGAGGTTTTTAAAGGGGGGCCTTTTGGTCTAACAATTTACCACAGGCTTAGAGTTCTGT
GATCAGTAGGCCAAATACCTAGTGAGGTC
TTGAGGAGGAAGAAGTAAAGGGAATCTGTTTACATTTATTTATTTGTTGTACATTACATATGG
GCACACTTGAGCCTTGGGCATGTGTGGAAGTTATAGGACAAACTGCTGGAATCCGTTAGCTCCT
AACATGTGGGATCCCGGATCGAGCTCAGGTCGTGAGGACTGACAGCAAGTGCCTTTACCCAT
CCGTTGTCCAGTCGCTGAGTCATCTTGGGGTCAGGCTTTTTCCCTTCCGCAGGGTTAGGTCA
TTGTTCTCTCAGTACATGTTTTCCCGACTTCTGTTAGAGGCCTTGAAGGAGGAGGATCCTGG
GAAATCTGAGGTCAGACATTTGAGCAATTCAGAGAATTCGGGAGGCTGTAAGGTAGAGAGAAA
GGACTTTGGGTGAGCTACAATGTCTTGAGGAAGTATTGTGTCACCTCCGA**CCCTC**AGTTTCCCT
GGGTGTCAAGTGCTCCTTTGTGCAACGTGTACGCTCCAGAGGAAACCGACTTATTCAACAG
GTGGTAATCCCAAGGTGAGAAAAGCAGACTCAACCGGAACAGCCAGGACAAAGACAATTCCC
CGGGGCGGATCCCGGATCGGTTT**CTCCT**AGGGCTGTGGCGAGGATGATTAGCAGCGGCACG
GGCACCTTTGGCCCGCACCTGGGCAGCTGGAATCCTCGGGGACGTGTGGGCCGGCGCGAG
CGGAGCTAGGGCGCGGGAAGGGGGCGGGCCTGGGCCGGTGGGCGGAGCGG

Smyd3: Binds DNA containing 5-CCCTCC-3 or 5-GAGGGG-3 sequences
5-GGAGGG-3or 5-CCCCTC-3

Supplementary Figure 4. The promoter region (2 kilobase upstream of transcription start site, or TSS) of PARP16 contains 6 potential Smyd3 binding sites



Supplementary Figure 5. EGCG decreases vascular inflammation markers in Ang II-infusion mice. Western blot of VCAM-1, iNOS, and GAPDH in aortic great vessels from mouse without Ang II infusion (control), Ang II-infused mouse and Ang II-infused mouse treated by 20, 50 mg/kg/day EGCG. ^{###} $p < 0.001$ vs. control; ^{***} $p < 0.001$ vs. Ang II group; all data were shown as mean \pm S.D, $n=6$ /group.



Supplementary Figure 6. PARP16 upregulation exists also in cellular replicated senescence models. (A) PARP16 upregulation exists in cellular replicated senescence models. Western blot of PARP16, senescence marker (p21), and SASP markers (VCAM-1, iNOS) in passage two (p2), and passage ten (p10) of replicative RAEC cells; (B) ER stress markers (p-eIF2 α , p-IRE α , p-PERK, cleaved ATF6, Bip and Spliced XBP-1) expression were determined by Western blot in passage two (p2) and passage ten (p10) of replicative RAEC cells.